



eDNA Pilot Project

AT LYNN HEADWATERS REGIONAL PARK

Robyn Worcester

NATURAL RESOURCES MANAGEMENT SPECIALIST

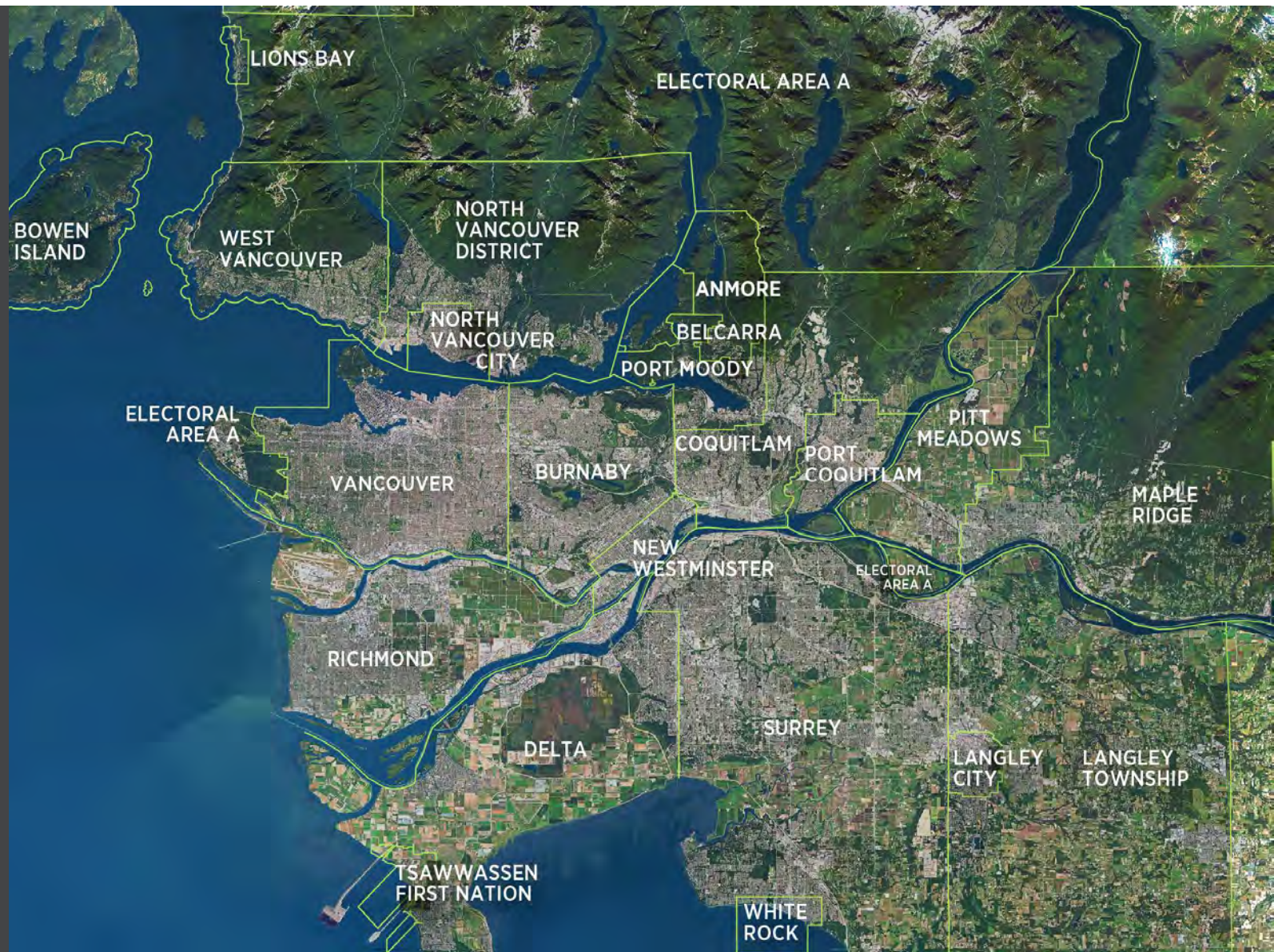
SCCP Conservation Connections, Oct 16, 2019

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Metro Vancouver

21 Municipalities,
1 Electoral Area,
and 1 Treaty
First Nation

WORKING TOGETHER FOR
A LIVABLE REGION



2 Ecological Conservancy Areas





OUR ROLE:
PROTECTING
Metro Vancouver's
natural areas
+
CONNECTING
people with them

What is eDNA?

- Genetic materials shed into the environment
- Most commonly used for species that use aquatic environments



Why use eDNA?

- Faster, easier, and cheaper than traditional techniques
- No animal interference required
- Helps with cryptic species
- No permits required



When to use eDNA?

- Consider species life cycle / timing
- Consider habitat - streams / rivers are most reliable
- When no abundance information is needed



How does the sampling work?

- Plan sampling sites
- Collect water samples
- Filter water to collect eDNA
- Send filters to the lab





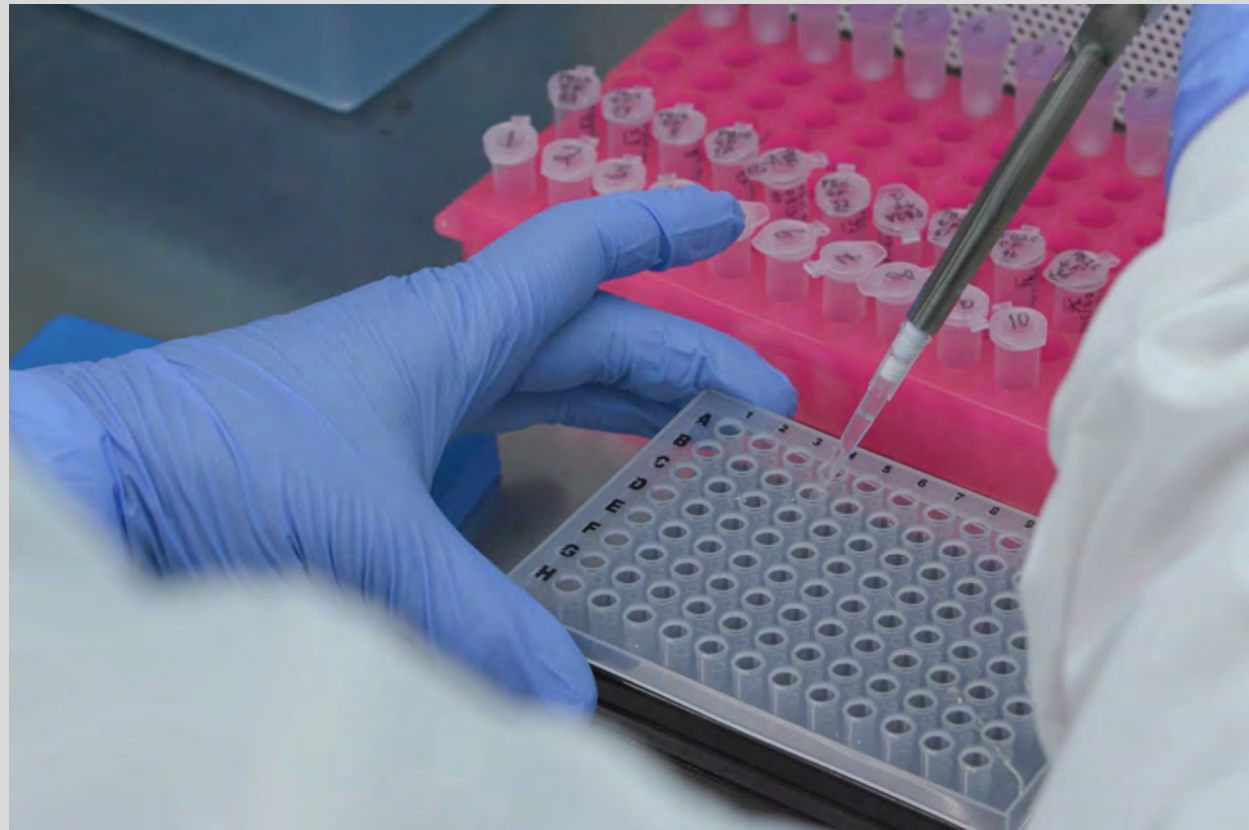
How does the filtering work?

- Water through special filters
- Filtration with a vacuum pump
- Filter dehydrated with self-indicating silica beads



What happens at the lab?

- ‘Clean’ samples that may need it
- DNA extracted and reviewed against primers developed for target species
- qPCR technique used



Other considerations?

- Paying attention to details in field sampling and lab filtration is key to reduce contamination
- Selecting sites, timing, and weather conditions matter

Environmental DNA Protocol for Freshwater Aquatic Ecosystems Version 2.2

Prepared for:
BC Ministry of Environment
Ecosystems Branch
2975 Jutland Road
Victoria, BC V8T 5J9

Prepared by:
Hemmera Envirochem Inc.
Jared Hobbs, M.Sc., R.P.Bio.
18th Floor, 4730 Kingsway
Burnaby, BC V5H 0C6

Washington State University
Dr. Caren Goldberg
100 Dairy Road, 404 Heald Hall
Pullman, WA 99164

With contributions and review from:
University of Victoria
Drs. Caren C. Helbing and Nik Veldhoen
Department of Biochemistry & Microbiology
Victoria, BC V8P 5C2

November 2017

Lynn Headwaters Regional Park Pilot

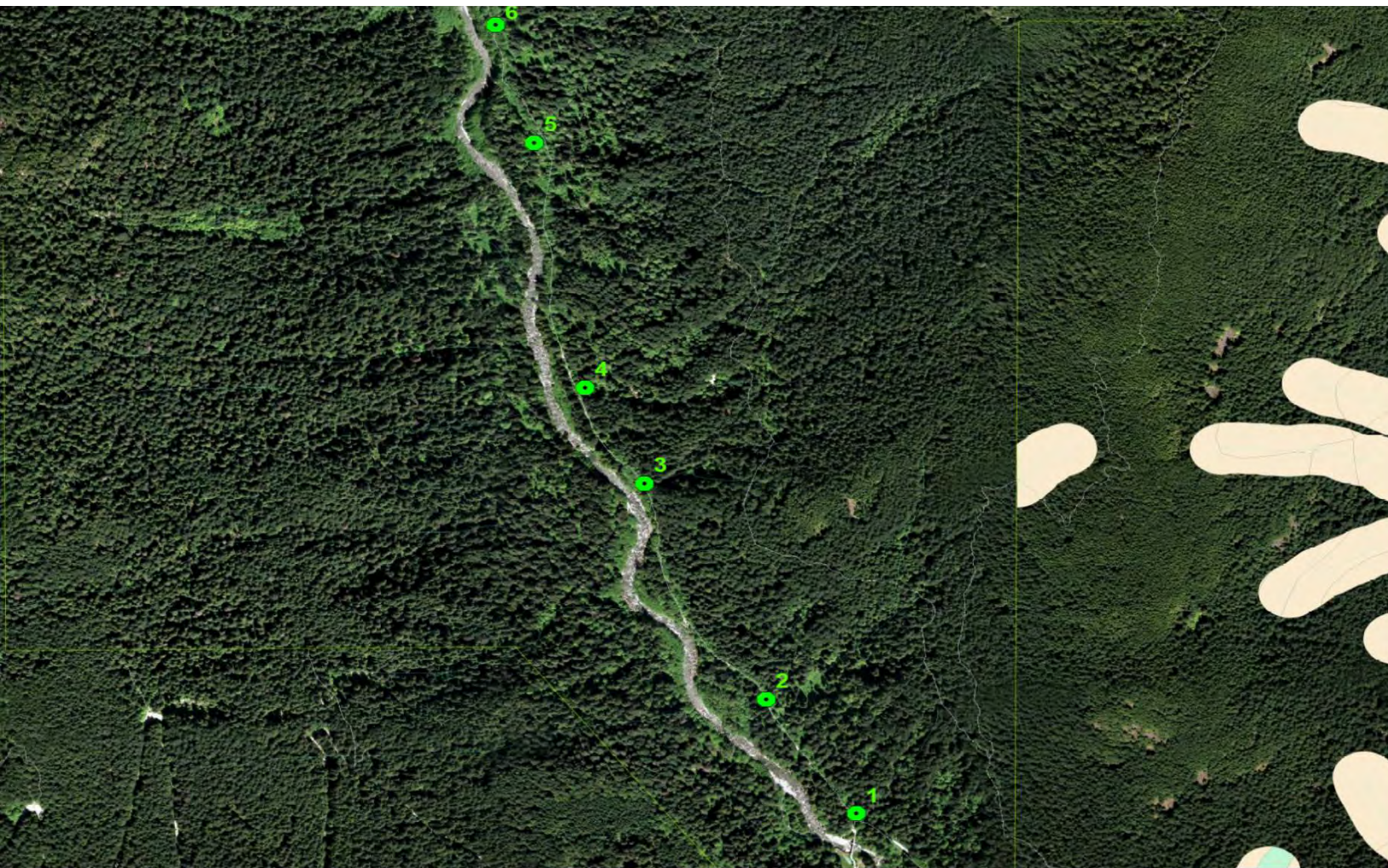
- Tested for 3 species during overlap of breeding season
- University of Victoria lab processed samples
- Film crew followed us around



Methods

- 6 sites (3 each) + one control sample = 19 samples
- DIY lab to process
- DRAFT RISC standards used





Legend

● eDNA Sampling Proposed

Trail and Road Network

Watercourse

<all other values>

WatercourseType

Canal

Dam or Indefinite Storage

Detention Pond Top

Flume

Hidden Drainage

Island

Pipeline

Rapids

River Stream

River Stream Banks

River Stream Indefinite

River Stream Intermittent

Sand Gravel Bar

Slough

Pacific Water Shrew

Coastal Tailed Frog - Species at Risk

Red Legged Frog




0 60 120 240 360 480 Meters

Results

qPCR Result (Lab Interpretation)

qPCR assay replicate (50 cycles/run)

Positive 


Negative 


Water sample replicate


Positive 

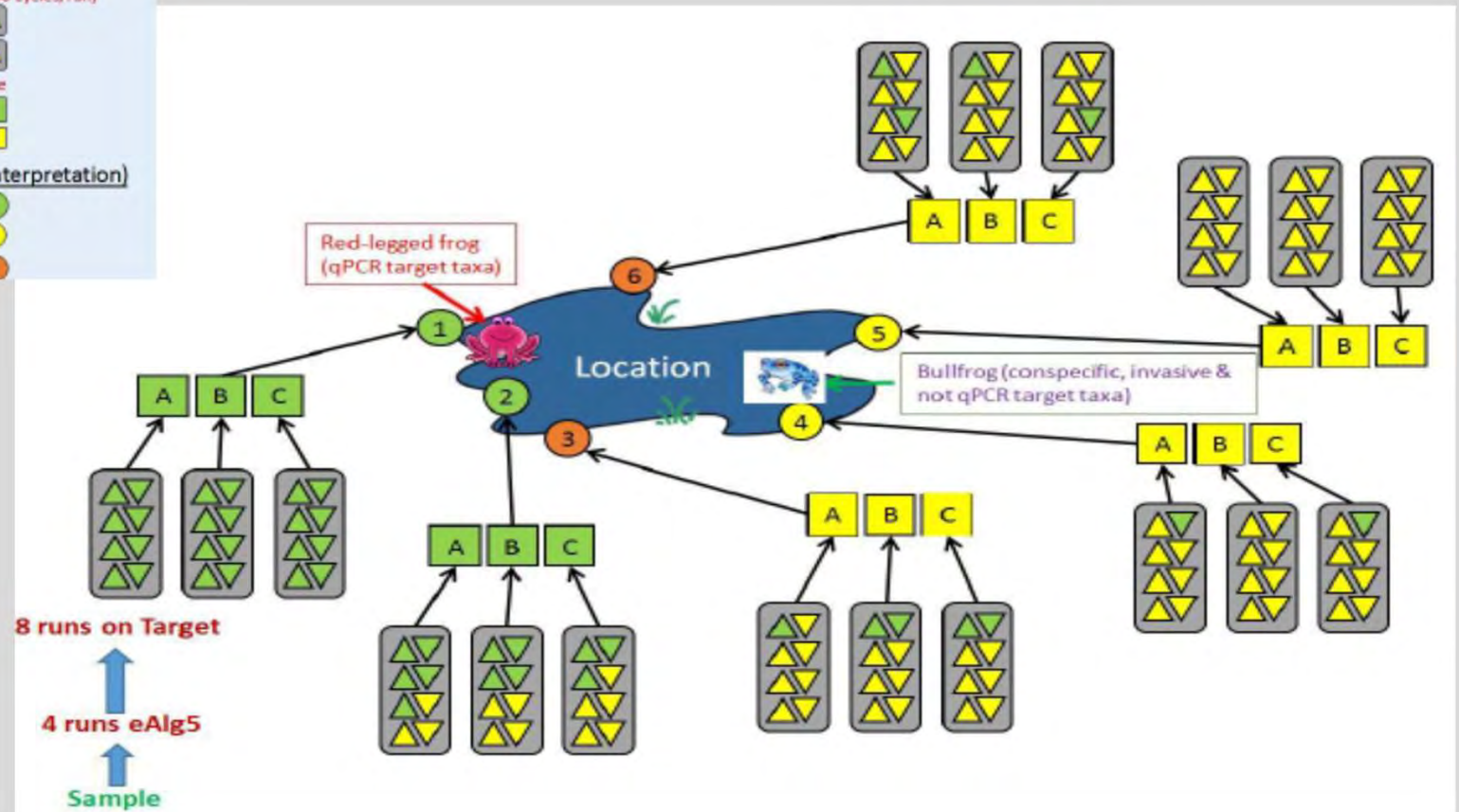
Negative 

Site (Biological Interpretation)

Positive 

Negative 

Suspected 



(Taken from NRTG training notes)

Sample replicate	Collection date	Test for (TT)	eASTR4 Frequency	Lab Call	Biol Call	eRAAU1 Frequency	Lab Call	Biol Call	eSOBE4 Re-extracted	Lab
LYN1-1	14-Jun-19	ASTR,RAAU,SOBE	8/8	Y	PI	2/8	Y	PI	0/8	M
LYN1-2	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	1/8	Y	PI	0/8	M
LYN1-3	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	3/8	Y	Y	0/8	M
LYN2-1	14-Jun-19	ASTR,RAAU,SOBE	5/8	Y	PI	0/8	N	PI	0/8	M
LYN2-2	14-Jun-19	ASTR,RAAU,SOBE	8/8	Y	PI	0/8	N	PI	0/8	M
LYN2-3	14-Jun-19	ASTR,RAAU,SOBE	6/8	Y	PI	2/8	Y	PI	0/8	M
LYN3-1	14-Jun-19	ASTR,RAAU,SOBE	5/8	Y	PI	0/8	N	PI	0/8	M
LYN3-2	14-Jun-19	ASTR,RAAU,SOBE	6/8	Y	PI	0/8	N	PI	0/8	M
LYN3-3	14-Jun-19	ASTR,RAAU,SOBE	7/8	Y	PI	0/8	N	PI	0/8	M
LYN4-1	14-Jun-19	ASTR,RAAU,SOBE	8/8	Y	PI	0/8	N	PI	0/8	M
LYN4-2	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	0/8	N	PI	0/8	M
LYN4-3	14-Jun-19	ASTR,RAAU,SOBE	8/8	Y	PI	1/8	Y	PI	0/8	M
1 of 1 LYN5-1	14-Jun-19	ASTR	0/8	N	PI	0/8	N	-	0/8	M
LYN5-2	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	4/8	Y	PI	0/8	M
LYN5-3	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	6/8	Y	PI	0/8	M
LYN6-1	14-Jun-19	ASTR,RAAU,SOBE	0/8	N	PI	8/8	Y	PI	0/8	M
LYN6-2	14-Jun-19	ASTR,RAAU,SOBE	1/8	Y	PI	0/8	N	PI	0/8	M
LYN6-3	14-Jun-19	ASTR,RAAU,SOBE	4/8	Y	PI	0/8	N	PI	0/8	M
	14-Jun-19	ASTR,RAAU,SOBE	6/8	Y	PI	0/8	N	PI	0/8	M

Results

- Tailed frogs in most of the fast streams
- Red-legged frogs in the low energy areas
- No Pacific Water Shrew detected






Future Projects

- Grant funding
- New primers

Learn more

- Training through Natural Resources Training Group
- Provincial standards developed by Hemmera (published in PLOS One)




RESEARCH ARTICLE

Implementation of Novel Design Features for qPCR-Based eDNA Assessment

Nik Veldhoen¹, Jared Hobbs², Georgios Ikononou¹, Michael Hii¹, Mary Lesperance³, Caren C. Helbing^{1*}

¹ Department of Biochemistry and Microbiology, University of Victoria, P.O. Box 3055, STN CSC, Victoria, British Columbia, V8W 2Y2, Canada, ² Hemmera Envirochem Inc., 303–1221 Broad Street, Victoria, British Columbia, V8W 2A4, Canada, ³ Department of Mathematics and Statistics, 3800 Finnerty Road, University of Victoria, Victoria, British Columbia, V8P 5C2, Canada

* chelbing@uvic.ca



Abstract

Environmental stewardship requires timely, accurate information related to the status of a given ecosystem and the species that occupy it. Recent advances in the application of the highly sensitive real-time quantitative polymerase chain reaction (qPCR) towards identification of constituents within environmental DNA (eDNA) now allow targeted detection of the presence of species-specific biological material within a localized geographic region. However, as with all molecular techniques predicated on the specificity and sensitivity of the PCR assay, careful validation of each eDNA qPCR assay in development must be per-

OPEN ACCESS

Citation: Veldhoen N, Hobbs J, Ikononou G, Hii M, Lesperance M, Helbing CC (2016) Implementation of Novel Design Features for qPCR-Based eDNA Assessment. PLoS ONE 11(11): e0164907. doi:10.1371/journal.pone.0164907



Questions?

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